**Laboratory #8**

**Chem 6614 Instrumental Methods of Chemistry**

**SUNY Alfred State College**

 **Gas Chromatography/Mass Spectroscopy: Part 2**

**Drug Bust Scenario**

*Lab notebook due Thursday 12 April*

**8.1. Background**

 Several different, suspicious powders found in unmarked plastic bags have been seized by US Border Protection officers during a search of Harry Hippie’s 1964 Volkswagen bus. Harry, who was attempting to cross back into the United States from Ciudad Juarez is a 59 year old male of indeterminate Caucasian ancestry whose passport shows an unusually large number of US-Mexico border crossings and who was-at the time of his detention- in possession of four rolls of twenty dollar bills totaling $4600. His appearance is dominated by an uncombed mane of graying shoulder-length hair and a psychedelic orange `Power to the People ’ tee shirt purchased at Jimmy Hendrix’s1970 Isle of Wight concert. Harry is currently a `guest’ in the DEA’s detention center near Allenby, Texas until the nature of the powders can be ascertained. Harry, unsurprisingly, swears that the powders are legit and that he is a victim of the Amerikan police state. As a side note, none of this would have happened had Harry simply refrained from calling the Border Protection Officer who requested his passport a `f…ing, fat, facist pig.’ Making `oink’ noises when asked to step out of his vehicle didn’t help either.

 **8.2. Purpose**

You are each drug analysts at the DEA who have been assigned a bag from Harry’s stash. You are tasked with doing the following

1. Prepare an initial elution of the sample powder in the bag.
2. Carry out an initial GC-MS examination on the initial elution.
3. If necessary, alter either the elution conditions or the GC-MS run conditions to obtain reasonable (SSS) peaks.
4. Provide an identification of the likely `drug’ (or drugs) -if any- in the elution using the MS Chem Station’s NIST search library.
5. Determine if Harry is in possession of a controlled substance, or whether he’s just a closet hypochondriac. You may need to use the Internet to check for common names of the compounds you identify in order to do this.

**8.3. Procedure**

**8.3.1. Initial elution**

Gently crush some of the powdered sample in a clean mortar and pestle and deliver a small spatula tip’s worth of the pulverized result to ~2 mL of dry methanol held in a small test tube.

Make sure to observe the approximate size of your spatula dose since you may have to re-make the elution again at higher or lower concentrations. Use a vortex mixer to `shake’ the sample and aid its dissolution. Allow the elution to stand and settle for a few minutes until the upper layer is clear. Extract the upper clear layer and store it in a clean, capped and labeled vial.

8.3.2 Initial GC-MS run

The Agilent instrument will be pumped down and ready the day of the lab. Because you are dealing with true unknowns a wide temperature range should be ramped over. To start it is suggested that an initial temperature of ~ 75 C and an upper temperature of 300 C be applied. The oven rate is your choice, but faster is better to start. Also, the parent masses of drugs can be quite high so the initial m/e window should be in the 40-400 amu range. The solvent delay may need to be adjusted (methanol boils at 65 C but it still can be resident after several minutes at 75) An initial starting solvent delay of 2 minutes is reasonable. To start, wet needle inject and a split ratio of 100:1 should be used. These can be adjusted if signal sizes are either too big or too small.

You must follow your sample run with a blank run (pure methanol) to purge the column of any residual using the same schedule as you applied to the sample.

8.3.3. Further GC-MS runs

While it is possible that the above conditions suffice to produce reasonable peak separations, you may need to adjust conditions. Since only one GC-MS instrument is currently available, you will need to allow other analysts to carry out their own initial runs before trying an amended run. For this reason, the 5890N Agilent GC-MS will be left on all week. You will need to schedule times with the instructor during the week to come in and use the instrument. Make sure to run a blank between sample runs or the column will become cluttered with residual peaks from previous runs.

You will also need to come in to use the Chem Station Data MS analysis software to search and identify your peaks likely chemical composition.

**8.4. Data and Observations**

As in experiment 7 the following instrument conditions for your unknown analysis should be recorded In **Data & Observations** :

**Table 1:**

**Method Parameters Used to Analyze Unknown # \_\_\_\_\_ on Alfred State Agilent 5975C GC/MS**

0) unknown # (in title)

1) secondary He tank pressure (in psi)

2) He flow rate

3) split ratio

4) auxiliary heater (transfer line) temperature

5) thermal ramp schedule

start T, hold time, ramp rate, end T, hold time, post-run time, equilibration time)

6) solvent delay time

7) injected sample volume

8) ambient, pre-injection manifold pressure in torr

9) quadrupole temperature

10) EI (source) temperature

11) mass range scanned (low to high)

12) file location for your Method

13) file location for your collected Data

You should also collect and place in **Data & Observations** a hard copy of the total ion chromatogram as well as summary mass spectral peak reports for ion peaks observed in the former.

Finally, make sure to describe how you prepared the sample you used in the first place.

**8.4.2 GC ion chromatogram**

The resulting chromatographic data should be re-tabulated as shown below.

**Table 2: Retention times, ion count and any comments vs component for unknown # \_\_\_\_ 1**

|  |  |  |  |
| --- | --- | --- | --- |
| **Component #** | **~ ion count** | **tr(minutes)2** | **Comments** |
| **1** | **23,000** | **1.9** | **Barely ahead of MeOH carrier** |
| **2** | **Etc** | **Etc** | **etc** |
| **3** | **Etc** | **Etc** | **etc** |

1include filename and address for total ion scan 2 peak retention time

**8.4.3 Component Mass Spectrum**

**Tables 3, 4,…:**

Tabulate as shown below, the 8-10 peaks with the largest % intensities for each significant component detected by GC.

**Table 3: major mass fragments observed for Component #1 (an example)**

 (see file.. include file name(s) and address for data and any analysis you do)

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|  |  |  |
| --- | --- | --- |
| m/e | Raw Peak height (or rel % = P/Pmax) | Comments |
| 42 | 28000 (28) |  |
| 77 | 40000 (40) |  |
| 82 | 100000 (100) | Pmax  |
| 96 | 35000 (35) |  |
| 105 | 30000 (30) |  |
| 182 | 70000 (70) |  |
| 198 | 10000 (10) |  |
| 272 | 10000 (10) |  |
| 303 | 15000 (15) |  |

**8.5 Calculations and Analysis**

 You will rely primarily on the suggested possible identities of the mass spectral peaks deduced by the NIST library assignment. Some common sense needs to be applied based on the case. Once you decide on the unknown’s component identities, you need to follow the format for assignment below . There will be no need to attempt fragment assignment since drug structure is too complex to allow unequivocal identification of the m/e peaks.

Components Assigned to Unknown # \_\_2\_\_\_ Based on GC/MS Analysis

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Component #** | **tr(min)** | **Ion count** | **Chemical identity** **(structural formula and name)** | **Notes** |
| **1** | **4.3** | 129000 | cocaine | Major component…this is pure dope ! |
| 2 | 5.1 | 13,000 | benzocaine | impurity |
| 3 | Etc | Etc | **Etc** |  |
|  |  |  |  |  |

Suggested Unknown Component 1 vs NIST1 reference peaks for Cocaine

 Unknown m/e rel intensity % NIST m/e for cocaine rel intensity %

 42 28 42 30

 77 40 77 38

 82 100 82 100

 96 35 96 41

 105 30 105 25

 182 70 182 74

 198 10 198 9

 272 10 272 11

 303 15 303(parent) 10

1 reference source

Suggested Unknown Component 2 vs NIST2 reference peaks for benzocaine

 Unknown m/e rel intensity % NIST m/e for benzocaine rel intensity %

Etc etc etc etc

2reference source

**Conclusions:**

Based on gc-ms analysis Unknown 2 contains DL cocaine (major) and benzocaine (minor).

(If other gc peaks were observed but weak, you should say something like:

“GC peaks at 2.5, 3.6 and 8 minutes were observed but at ion counts ~ 1% of those seen for the two components assigned above.”